Plant assemblage from the Upper Miocene deposits of the Belchatów Lignite Mine (Central Poland)

GRZEGORZ WOROBIEC1, ELŻBIETA WOROBIEC1 and ADAM SZYNKIEWICZ2

1 Władysław Szafer Institute of Botany, Polish Academy of Sciences, Lubićz 46, 31-512 Kraków, Poland; e-mail: g.worobiec@botany.pl, e.worobiec@botany.pl
2 Institute of Geological Sciences, Wrocław University, Pl. M. Borna 9, 50-204 Wrocław, Poland; e-mail: aszyn@ing.uni.wroc.pl

Received 16 March 2012; accepted for publication 19 September 2012

ABSTRACT. The Upper Miocene plant macroremains (mainly leaves) collected in the Belchatów Lignite Mine (Central Poland) were investigated and supplemented with palynological analysis. In the fossil flora there were found macroremains of Acer, Betula, Carpinus, Dicotylophyllum, Fagus, Laria, Liquidambar, Populus, Pterocarya, Quercus, Reevesia, Salix, Taxodium, Trapa, and Ulmus. Four new fossil leaf species to the Polish tertiary were reported: Quercus roburoides Gaudin, Reevesia hurnikii Kvaček, Dicotylophyllum sp. 1, and Dicotylophyllum sp. 2. The fossil plant assemblage is composed of mesophytic (zonal) and riparian (azonal) elements. The floristic composition confirms the Late Miocene age of the clayey-sandy unit of the Belchatów Lignite Mine and suggests warm temperate climate conditions.

KEYWORDS: plant macroremains, macromorphology, plant cuticles, micromorphology, palynology, palaeovegetation, Late Miocene, Poland

INTRODUCTION

In the course of palaeobotanical survey in the outcrop of the Belchatów Lignite Mine in 2004, an assemblage of abundant accumulations of plant macroremains, mainly fossil leaves, have been found. Good state of preservation of plant fossils made possible an extensive study of plant macroremains combined with palynological analysis. The investigations were aimed at palaeofloristics and palaeoecology of the assemblage to document biodiversity of the Neogene flora and vegetation of the Belchatów Lignite Mine.

GEOLOGY

The Belchatów Lignite Mine is situated in Central Poland, in the region between the rivers Warta and Pilica, ca 15 km south of the town Belchatów (Fig. 1). Lignite seams occur within a tectonic depression named the Kleszczów Graben that is filled with Neogene and Quaternary deposits (Gotowała & Hałuszczak 1999). A lithostratigraphic profile of the Neogene deposits filling the Kleszczów Graben (Fig. 2) with four main lithological units: subcoal unit (PW), coal unit (W) including the main seam (PG), clayey-coal unit (I-W), and the youngest clayey-sandy unit (I-P) has been established (Stuchlik et al. 1990, Czarnecki et al. 1992, Szynkiewicz 2000, Matl 2000). Multi-proxy studies (geological, palaeobotanical and palaeozoological) of the Neogene deposits at the Belchatów Lignite Mine allowed determination of the age of the main coal seam (PG) as Early Miocene, and those of the overburden sedimentary series of the clayey-sandy unit as Late Miocene (Burchart 1985, Burchart et al. 1988, Stuchlik et al. 1990, Kowalski 1993, Kowalski & Rzebik-Kowalska 2002, Stuchlik and Szynkiewicz 1998, Szynkiewicz 2000).
Investigated plant assemblages were collected from the Bełchatów Lignite Mine outcrop. Material was found in July, 2004 on the western slope of the open pit of the Bełchatów Lignite Mine, on the overburden escarpment 3, near the conveyor belt B-301. Its stratigraphical position corresponds to the lower part of the clayey-sandy unit of Late Miocene age (Fig. 2).

MATERIAL AND METHODS

Fossil specimens were slowly dried and prepared in the laboratory at the Władysław Szafer Institute of Botany, Polish Academy of Sciences. Plant macroremains are preserved mostly as compressions, often fragmented and deformed. The state of preservation of remains is usually good allowing preparation of fossil cuticles, exceptionally even isolation of leaf blades (Pl. 2, fig. 6). However, most of leaf compressions were strongly cracked which made the isolation of cuticles very difficult, in many cases even impossible.

From the selected leaf compressions fragments of coalified leaf lamina were sampled for cuticular analysis. The laminar fragments were cleaned with hydrofluoric acid, washed in water, macerated using a NaClO solution (a commercial bleach, called “Bielnar”), and finally mounted on slides with glycerine jelly.

All studied fossil specimens are housed at the W. Szafer Institute of Botany Polish Academy of Sciences, Kraków under the catalogue number KRAM-P 250. The total number of specimens from all taxa is greater than the total number of deposit fragments with plant remains because on some specimens several taxa were preserved. In such cases alphanumeric specimen number is used. In total 159 deposit samples were investigated.

Leaf macromorphological descriptions follow mostly Ellis et al. (2009) and cuticular descriptions (micromorphology) follows Dilcher (1974) and Wilkinson (1979). The way of measuring the size of micromorphological structures depended on the shape of the object. The diameter was measured in regular, round, and broadly elliptic objects, but the length and the width were measured in all rectangular objects. In structures of irregular or variable shape (mainly unmodified epidermal cells) the longest dimension was measured.

The drawings were made using a Carl Zeiss stereomicroscope with camera lucida attachment. Macrophotographs were taken using a NIKON Coolpix 995 digital camera and NIKON SMZ 800 stereomicroscope fitted with NIKON DS-5M-U1 digital camera and microphotographs were taken using NIKON Eclipse E400 microscope fitted with CANON A640 digital camera.

Macromremains were identified on the basis of macrofossil morphology and cuticular analysis. Plant remains were compared to other related fossil taxa and herbarium material for more precise identifications. Cuticles from the fossil specimens were compared to cuticles of fossil and extant plants to further support the taxonomic affinity of the fossils. The reference herbarium and collections of epidermal preparations from fossil and recent leaves are housed in the Department of Palaeobotany, W. Szafer Institute of Botany Polish Academy of Sciences, Kraków.
Two samples from the sediments containing leaf macroremains were used for palynological study (dr. E. Worobiec). The samples were prepared following a modified Erdtman’s acetolysis method (Moore et al. 1991), using hydrofluoric acid to remove clay minerals. Two slides from each sample were prepared, using glycerine jelly as a mounting medium. The palynomorphs were identified using comparative material from previous publications (e.g. Stuchlík et al. 2001, 2002, 2009, in press) and the reference palynological collection of the Department of Palaeobotany, W. Szafer Institute of Botany Polish Academy of Sciences, Kraków.

RESULTS

SYSTEMATIC PALAEOBOTANY

The classification for gymnosperms follows Gadek et al. (2000). The classification of angiosperms and author names of families follows the APG III (2009). Plant families, genera, and species are listed alphabetically.

Gymnosperms

Cupressaceae s.l. sensu Gadek et al. 2000
Taxodioidae Endl. ex K. Koch sensu Gadek et al. 2000

Taxodium Rich.

**Taxodium dubium** (Sternberg) Heer

Pl. 1, fig. 5, Pl. 6, fig. 1

1823 *Phyllites dubius* Sternberg; Sternberg, p. 37, Pl. 36, fig. 3.
1853 *Taxodium dubium* (Sternberg) Heer; Heer, p. 136.
1855 *Taxodium dubium* (Sternberg) Heer; Heer, p. 49, Pl. 17, figs 5-15.


Description. Macromorphology. Fragments of leafy shoots, up to 5.3 cm long, vestigial needles at a base of shoot. Needles above base entire-margined, 0.7–1.2 cm long and 0.10–0.12 cm wide, decreasing in length towards base of shoot, apex of needleacute, rarely acuminate, base decurrent. Midvein clearly visible; epidermal cells mostly rectangular or square, 40–75 μm long and 20–32 μm wide. Cell walls thick, straight or rounded, end walls perpendicular or oblique to the long axis of cells. Hypodermis present, composed of cells of similar dimensions as epidermis and with pitted walls. Cuticle rather thick, slightly granulate, probably due to presence of epicuticular wax. Stomata irregularly-elliptic, 45–50 μm long. Long axis of stomata irregularly oriented, usually oblique to the long axis of needle and only sporadically parallel. Distinct T-shaped thickenings at the stomatal poles. Outer stomatal aperture elongate-elliptic, about 20 μm long and 5–7 μm wide.

Remarks. The *Taxodium* shoots are morphologically similar to those of *Sequoia abietina* (Brongniart) Knobloch, but differ from the latter by having slightly narrower and longer needles. However, the needle morphology of both *Sequoia* and *Taxodium* is highly variable. For a reliable identification, the epidermal micromorphology of the needles from Belchatów was analysed. Epidermis of *Taxodium* shows that the long axes of stomata are oblique to the axis of the needle. In case of taxodioid shoots of *Sequoia* the long axis of stomata is more or less parallel to the axis of the needle. In addition, the cuticle of *Taxodium* needles is rather delicate and was strongly damaged during preparation procedure, while the epidermis of *Sequoia abietina* needles is strongly cutinized and resistant to overmaceration.

Modern equivalents. *Taxodium dubium* is comparable with two extant species from North America: *T. distichum* (L.) Rich. and *T. mucronatum* Tenore (Hummel 1983). *Taxodium distichum* grows in swamps and on river banks in the south-western USA while *T. mucronatum* is associated with wet habitats on banks of mountain rivers and streams in Mexico (Krüssmann 1972).


Occurrence. *Taxodium dubium* was a common component of European tertiary azonal vegetation. It occurred from the Late Eocene to Late Miocene in the Atlantic- Boreal province sensu Mai of Central Europe and from the Late Oligocene to Late Pliocene in the Transeuropean Paratethys province sensu Mai.
Dicotyledones

Altingiaceae Horan., nom. cons.

**Liquidambar** L.

**Liquidambar europaea** A. Braun

Fig. 3: 1–2, Pl. 1, figs 1–3

1836 *Liquidambar europaeum* A. Braun in Buckland, p. 513.

1969 *Liquidambar europaea* A. Braun; Knobloch, p. 94, pl. 44, figs 1, 3, 4–7, pl. 45, figs 1, 2, 6, pl. 46, figs 1, 4, pl. 59, fig. 2.


**Description.** Macromorphology. Leaves palmately lobed, 3–5 lobed, up to 7.5 cm across. Apex of lobes attenuate. Leaf margin simple serrate, teeth small, apical side of teeth very short, concave, basal side long, acuminate, teeth apex round. Veins terminate in sinus between teeth. Primary venation palmate-actinodromous, perfect, primary vein branched into 5 branches. Secondary venation semicraspedodromous, secondaries curved upwards, interconnected near leaf margin, forming loops, sending of fine veins towards teeth. Tertiary veins percurrent or weakly percurrent.

**Remarks.** Shape of leaf, distinctive serration of margin and network of venation are characteristic for the genus *Liquidambar* L. and the fossil species *Liquidambar europaea* A. Braun with typically 5-lobate leaves.

**Modern equivalents.** According to Hummel (1983) extant species from the section *Euliquidambar* Harms (especially the North American *Liquidambar styraciflua* L. and *L. orientalis* Mill. from Asia Minor) are closely related to the fossil species *L. europaea*.


**Occurrence.** Common in the Neogene floras of Europe. From Poland it is known from the Middle Miocene to Pliocene (for localities see Hummel 1983).

**Betulaceae** Gray, nom. cons.

**Betula** L.

**Betula plioplatyptera** Hummel

Fig. 3: 4, Pl. 1, figs 4, 6

1991 *Betula plioplatyptera* Hummel, pp. 82–87, Fig. 6: 1–7, pl. 3, figs 1–3, pl. 4, figs 1–1a, pl. 5, figs 1–4.

**Material.** KRAM-P 250: 13/I, 83/I, 97/I. *Betula cf. plioplatyptera*: 156/I.

**Description.** Macromorphology. Leaf fragments, up to 5.5 cm long and 2.5–2.8 cm wide. Leaf apex acute, base acute and decurrent, usually without teeth. Leaf margin double serrate, apical and basal sides of teeth acuminate, sometimes rounded. Tooth apex mostly acute, sometimes slightly rounded. Secondary veins and subsidiary veins terminating at the teeth apex. Venation pinnate, primary vein straight, of moderate thickness. Secondary venation simple craspedodromous, preserved up to 8 pairs of secondaries, veins first straight curve upwards near leaf margin, distributed at intervals of 0.3 cm near leaf base to 0.4–0.7 cm in the middle of the lamina. Secondary veins diverging from mid-vein at an angle of 40–55°. Tertiary venation percurrent, ca. 8–10 tertiary veins occur per 1 cm of secondary vein length. Tertiary venation forms an angle of 120–135° with the primary vein.

**Remarks.** The macromorphological features of leaf fragments, e.g. shape of the leaves, large primary teeth, minor secondary teeth and an irregularly serrate leaf margin allowed its identification as *Betula plioplatyptera* Hummel. Specimen no. 156/I only presumably belongs to *Betula plioplatyptera* as it is badly preserved. This species differs from *Betula subpubescens* Goeppert emend. Worobiec in shape, outline of leaf base, arrangement of teeth, and course of the tertiary venation (Hummel 1991, Worobiec & Szynkiewicz 2007).

**Modern equivalents.** *Betula plioplatyptera* resembles the extant North American species *B. papyrifera* Marsh., particularly its form *elongata* Schneider though it is considerably...
different when regarding the structure of epidermis (Hummel 1991).

Ecology. Arctotertiary, temperate element. Most probably component of riparian or mesophytic forests.

Occurrence. In Poland it is found in Mioocene deposits of Belchatów (Worobiec & Szynkiewicz 2007) and Early Pliocene of Ruszów (Hummel 1991).

**Betula subpubescens** Goeppert emend. Worobiec

Fig. 3: 5, Pl. 1, fig. 7, Pl. 6, fig. 2

1855 *Betula subpubescens* Goeppert, p. 11, Pl. 3, fig. 9.
1991 *Betula subpubescens* Goeppert; Hummel, pp. 87–91, Pl. 6, figs 1–4, Pl. 7, figs 1–4, Pl. 8, figs 1–5; Fig. 7: 1–13.
2007 *Betula subpubescens* Goeppert emend. Worobiec, Worobiec & Szynkiewicz, pp. 43–45, Pl. 6, figs 2, 3a–3c, Pl. 7, figs 2a–2e.


Description. Macromorphology. Leaves elliptic to ovate, up to 5.0 cm long, 1.4–3.2 cm wide, leaf apex acute to acuminate, base obtuse, often decurrent. Petiole, rarely preserved, up to 1.4 cm long. Leaf margin double serrate, leaf base often entire-margined. Primary teeth served by secondary veins are large, subsidiary teeth (2–3) small, apical and basal sides of teeth usually acuminate, tooth apex acute or acuminate. Secondary veins and subsidiary veins terminate at apex of teeth. Venation pinnate, primary vein straight, of moderate thickness. Secondary venation simple craspedodromous, 8–11 pairs of secondaries preserved, arise at intervals of 0.2–0.3 cm near leaf base and 0.3–0.9 cm in middle of the lamina, initially ± straight and curved upwards near leaf margin, diverging from midvein at an angle of 55–60° near leaf base and 40–55° (commonly 40–50°) in the middle of the lamina. Tertiary venation percurrent.

Micromorphology. Adaxial epidermis composed of more or less isodiametric cells, 20–25 μm across with usually straight anticlinal cell walls. Leaves hypostomatic. There are only visible some outer stomatal ledge apertures, wide spindle-shaped, about 18 μm long and 10 μm wide. On the abaxial epidermis glandular trichome bases, multicellular (8–10 cells), 30–35 μm across were found.

Remarks. Macro- and micromorphologically the above described leaf remains correspond to leaves of *Betula subpubescens* Goeppert emend. Worobiec. Some specimens only presumably belong to *Betula subpubescens* as they are badly preserved.

Modern equivalents. *Betula subpubescens* is comparable to recent *Betula pumila* Ehrh. (= *Betula alba* Roth), *B. bhojpatra* Wall. (= *B. utilis* D.Don.) and *B. davurica* Pall. (Hummel, 1991), as well as according to Němejc (1949) to *B. verrucosa* Ehrh. (= *B. pendula* Roth).

Ecology. Arctotertiary, temperate element. Most probably a component of riparian or mesophytic forests.

Occurrence. In Poland it is found from the Middle Miocene to Early Pliocene (Krajewska 1998).

**Betula sp.**


Description. Macromorphology. Fragments of leaves with *Betula* type of teeth. Leaf margin double serrate, primary teeth served by secondary veins are large, apical and basal sides of teeth acuminate, apex of tooth acute or moderate acute.

Remarks. The leaves undoubtedly represent the genus *Betula*. Identification to species level is impossible due to the incompleteness of the specimens, but doubtless among them are represented species *Betula piolalptypeta* Hummel and *Betula subpubescens* Goeppert emend. Worobiec and perhaps also *Alnus adscendens* (Goeppert) Zastawniai & Walther.

**Carpinus L.**

*Carpinus grandis* Unger emend. Heer

Fig. 3: 7, Pl. 2, figs 2, 4a–4b

1850 *Carpinus grandis* Unger, p. 408.
1852 *Carpinus grandis* Unger, p. 39, Pl. 20, figs 4, 5.
1856 *Carpinus grandis* Unger; Heer, Pl. 71, figs 19b, c–e, Pl. 72, figs 2–11, 14, 16, 17, 19, 20, 22–24, Pl. 73, figs 2–4.


**Description. Macromorphology.** Leaf blade frequently characteristically wavy. Leaves elliptic or ovate, up to 5.0 cm long and 2.5–3.4 cm wide with acute, obtuse, rounded, or rarely slightly cordate base. Petiole up to 1.0 cm long. Leaf margin double serrate, teeth small, numerous, more or less of the same size, apical and basal side of teeth acuminate or rounded. Teeth apex acute, teeth often slightly hook-shaped. Secondary veins and their branches terminate at the teeth apex. Venation pinnate, primary vein straight of moderate thickness. Secondary venation simple craspedodromous. Secondaries preserved in up to 8 pairs, straight, near leaf margin slightly upward curved, distributed at intervals of 0.3–0.4 cm near leaf the base and 0.5–0.8 cm in the middle part of lamina. Secondary veins form an angle of 40–50° with the primary vein, close to the leaf margin produce numerous branches entering the nearest teeth. Tertiary venation percurrent, forms an angle of 120–140° with the primary vein. About 9 to 10 tertiary veins occur per 1 cm of secondary vein length.

**Remarks.** The elliptic shape, double serrate leaf margin, arrangement of secondary veins, and venation of the leaf base are characteristic of the genus *Carpinus* L. The leaves under discussion belong to the fossil species *Carpinus grandis* Unger emend. Heer, most probably a collective species that includes several morphotypes of fossil leaves (Mai & Walther 1978, 1988, Hummel 1991, Zastawniak & Walther 1998).


**Ecology.** Arctotertiary, temperate element. Component of mesophytic and probably also riparian vegetation.

1850 *Carpinus grandis* Unger, p. 408.

1953 *Carpinus grandis* Unger sensu Berger; Berger, p. 14, Figs 5: A–G.

**Material.** KRAM-P 250: 5/II, 53/III, 54/IV, 122/II.

**Description.** Fragments of fruit involucre.

**Remarks.** These remains of fruits of fossil hornbeam represent species *Carpinus grandis* Unger sensu Berger compared with involucres of recent *Carpinus betulus* L. (Krajewska 1998).

**Betulaceae indet.**


**Remarks.** On the basis of the type of serration of leaf margins, these fragments of leaves could be categorised as belonging undoubtedly to the family Betulaceae. Due to poor state of preservation identification of the genera position is uncertain.

**Dicotyledones incertae sedis**

**Dicotylophyllum sp. 1**


**Description. Macromorphology.** Fragments of entire-margined leaves, up to 4.5 cm long and 1.4–2.6 (3.6) cm wide. Leaf apex probably attenuate, leaf base obtuse. Venation pinnate, primary vein straight, and of moderate thickness. Secondary venation brochiodromous. Secondaries upwardly curved and interconnected in loops, arranged alternately,
distributed at intervals of 0.5–1.0 (commonly 0.6–0.8) cm. Secondary veins form an angle of 50–70° with primary vein. Tertiary venation reticulate. Higher order venation random reticulate. Areoles well developed, 0.20–0.45 mm across, veinlets usually simple, rarely once branched or lacking. Marginal ultimate venation looped.

**Micromorphology.** Cells of abaxial epidermis with usually straight, rarely rounded walls, about 20 μm across. Leaves hypostomatic. Stomata anomocytic, narrow elliptic to elliptic, 15–25 μm across. Outer stomatal ledge aperture spindle-shaped, usually rather narrow, 9–17 μm long and 2.5–6.0 μm wide. On the abaxial epidermis, unicellular, strongly cutinised, rounded to elliptic trichome bases were found, 12–22 μm in diameter. Trichome bases are located both over the veins and on vein-free area.

**Remarks.** Systematic position of the described leaf fragment is unknown. Some similarities in macromorphology could be observed in comparison with leaves described by Kvaček et al. (2011) as species of morphogenus *Leguminosites* Bowerbank emend. Schimper, especially with *Leguminosites* sp. A. However, they differ from *Dicotylophyllum* sp. from Belchatów in the development of veinlets and micromorphology of epidermis.

**Dicotylophyllum sp. 2**

*Fagus l.*

**Fagus silesiaca** Walther & Zastawniak

1991 *Fagus silesiaca* Walther & Zastawniak, p. 156–160, Fig. 1, pl. 1, figs 1–6, pl. 2, fig. 1.


**Description.** Macromorphology. Leaves usually elliptic, rarely ovate, up to 6.0 cm long and 2.6–3.8 (commonly 3.0–3.5) cm wide with acute or obtuse, exceptionally even asymmetric leaf base. Leaf margin simple serrate, teeth upwardly curved, apical sides of teeth concave, basal sides acuminate or convex, teeth apex acute or slightly rounded. Venation pinnate, primary vein in apical part of leaf often characteristically undulate. Secondary venation simple craspedodromous, secondaries preserved in 8–10 pairs, straight or near leaf margin slightly upwardly curved, distributed at intervals of 0.5–1.0 (commonly 0.6–0.8) cm. Secondary veins form an angle of (35) 40–50 (55)° with primary vein. Tertiary venation percurrent, forms an angle of 120–140° with the primary vein. Tertiary veins occur per 1 cm of secondary vein length. Higher order venation orthogonal reticulate. Areoles well developed, 0.3–0.5 mm across, veinlets simple or none. Marginal ultimate venation looped.

**Micromorphology.** Adaxial epidermis composed of usually slightly elongated cells with
distinctly undulate cell walls, about 37 μm across. Leaves hypostomatic. Abaxial epidermis consists of variable-shaped cells with rounded or slightly undulate cell walls, 25–30 μm across. Stomata usually roundish, rarely wide elliptic, 20–25 μm in diameter. Outer stomatal ledge elliptic-oblongate, 7–11 μm long and 5–7 μm wide. Indistinct polar cuticular thickness usually visible.

**Remarks.** The fragments of fossil beech leaves described above belong to *Fagus silesiaca* Walther & Zastawniak. It differs from the leaves of *Fagus kraeuselii* Z. Kvaček & Walther in having a greater number of secondary veins (8–10 pairs) and from middle Oligocene – Early Miocene species *Fagus saxonica* Z. Kvaček & Walther in having fewer secondary veins (comp. Kvaček & Walther 1991). Rather similar to that of *Fagus silesiaca* is *Fagus menzelii* Z. Kvaček & Walther (Kvaček & Walther 1991). The present author confirms his earlier opinion (Worobiec 2003) that there are no significant taxonomic differences between *Fagus silesiaca* and *F. menzelii* and both morphotypes most probably represent one taxon. As the description of *Fagus silesiaca* is earlier, the name *Fagus silesiaca* has been given preference, in accordance with the International Code of Botanical Nomenclature. Discussed leaves of *Fagus silesiaca* Walther & Zastawniak differ from Neogene Iceland species *F. friedrichii* Grimsson & Denk having less number of secondary veins and form *F. gussonii* Massalongo in generally smaller leaves (Grimsson & Denk 2005).

**Modern equivalents.** The macromorphology and micromorphology of leaves of *Fagus silesiaca* is comparable to that of the recent *Fagus hayatae* Palibin ex Hayata that grows in the Mixed Mesophytic Forests of central China (Dyjor et al. 1992). *Fagus grandiflora* Ehrh. (= *Fagus ferruginea* Ait.) from the eastern USA is morphologically very similar to *Fagus silesiaca*, but differs in having anomocytic stomata (Kvaček & Walther 1991).

**Ecology.** Arctotertiary, warm temperate element. According to Kvaček and Walther (1991) it grew in mixed mesophytic or deciduous forests.

**Occurrence.** *Fagus silesiaca* is frequently found from the Late Miocene to the Early Pliocene of Central Europe (Walther 1994) and was usually reported under the incorrect names *Fagus attenuata* Goeppert and *Fagus haidingeri* Kováts sensu Knobloch. In the territory of Poland found from Middle Miocene to Pliocene (see Worobiec 2003).

**Quercus L.**

**Quercus pseudocastanea** Goeppert emend. Walther & Zastawniak

Pl. 3, fgs 2a–2b

1991 *Quercus pseudocastanea* Goeppert emend. Walther & Zastawniak, p. 169, Fig. 8, pl. 2, fgs 2–6, pl. 3, fgs 1–6.


**Description.** Macromorphology. Usually small fragments of leaves (up to 5 cm). Leaf margin lobate, lobes 0.8 cm high and up to 1.2 cm in diameter in basal part, apical and basal sides of lobes convex, lobes apex always acute. Secondary veins terminate at the lobes apex. Venation pinnate, secondary venation on preserved fragments simple craspedodromous. Secondary veins diverge from the main vein at an angle of about 50° at intervals of 0.9–1.0 cm. Tertiary venation percurrent and forms an angle of about 135° with the primary vein.

**Remarks.** The shape of the leaf fragments and the marginal lobes with always acute apex are characteristic of the species *Quercus pseudocastanea* Goeppert emend. Walther & Zastawniak. *Quercus gigas* Goeppert & Zastawniak differs markedly by having serrate margin with spinose teeth apex.

**Modern equivalents.** With respect to the macro- and micromorphology *Quercus pseudocastanea* resembles recent oaks from section *Cerris* Oersted (Walther & Zastawniak 1991).

**Ecology.** Arctotertiary, warm temperate element. Most probably component of mesophytic forests (Belz & Mosbrugger 1994).

**Occurrence.** *Quercus pseudocastanea* is common in the tertiary of Europe. It is found from the Middle Miocene to the Upper Pliocene. In the territory of Poland found from the Middle Miocene to Pliocene (see Walther & Zastawniak 1991).
Quercus roburoides Gaudin

Pl. 3, figs 1a–1b

1859 Quercus roburoides Gaudin; Gaudin & Strozzi, p. 44, pl. 3, fig. 14.


Description. Macromorphology. Fragments of leaves up to 8.0 cm long and presumably 8.0 cm wide, leaf base rounded. Leaf margin lobate, lobes 0.3 to 1.3 cm high and 0.8 to 2.0 cm in diameter in basal part, apical and basal sides of lobes convex, lobes apex always rounded, often retuse (with apical cleft). Secondary veins terminate at the lobes apex. Venation pinnate, secondary venation on preserved fragments simple craspedodromous. Secondary veins more or less straight, irregularly spaced at intervals of 0.5–2.0 cm, diverge from the main vein at an angle of 30–60°, the angle of divergence of the secondary veins considerably decrease from base (60°) to apex (30°). Tertiary venation percurrent, on one specimen about 7 tertiary veins occur per 1 cm of secondary vein length.

Micromorphology. On the unsuccessful cuticular slide there are only visible scattered stomata, 17–25 μm across with T-shaped thickenings at the stomatal poles.

Remarks. These leaves are similar to leaves of the fossil oak Quercus pseudocastanea Goeppert emend. Walther & Zastawniak. However, some differences in macromorphology between them could be found. Leaves of Quercus pseudocastanea from the same locality are rather regular in shape and always have lobes with acute apex. The discussed leaves are usually of less regular shape and always have lobes with rounded, often even retuse apex. This last feature concerning lobes is according to Belz and Mosbrugger (1994) typical for Quercus roburoides Gaudin. Detailed description and comparison of macro- and micromorphology of leaves of both Quercus pseudocastanea and Quercus roburoides was made by Van der Burgh (1993). In his opinion Quercus pseudocastanea has regular shape, numerous secondary veins with almost absent intersecondaries and smaller lobes contrary to Quercus roburoides which leaves are more irregular in shape and less number of lobes. Van der Burgh (op. cit.) has not found any differences in the micromorphology of cuticles between these two discussed species.

Modern equivalents. In the respect of macromorphology, recent oaks Quercus petraea (Matt.) Liebl. and Quercus canariensis Willd., members of the section Robur are most similar to Quercus roburoides (Belz & Mosbrugger 1994).


Occurrence. Taxon new for the fossil floras of Poland. Leaves reported as Quercus pseudorobur Kovats (should be treated as a synonym of Quercus roburoides) from the Middle Miocene (Sarmatian) locality of Młyny (Zastawniak 1980) belong to species Quercus pseudocastanea (Walther & Zastawniak 1991). Quercus roburoides is a typical species of the late Neogene floras of Europe, found in the Pliocene deposits (Van der Burgh 1993, Walther 1994).

Quercus sp.


Remarks. Leaves macromorphologically similar both to Quercus pseudocastanea Goeppert emend. Walther & Zastawniak and Quercus roburoides Gaudin, but too poorly preserved for unambiguous identification.

Juglandaceae DC. ex Perleb, nom. cons.

Pterocarya Kunth.

Pterocarya paradisiaca (Unger) Ilinskaya

Fig. 4: 3, Pl. 3, figs 6a–6b, Pl. 6, figs 6a–6b

1849 Prunus paradisiaca Unger, p. 7, pl. 14, fig. 22.
1897 Pterocarya castaneifolia (Goeppert) Schlechtendal; Schlechtendal, p. 22, pl. 5, figs 1–3b, pl. 6, figs 5–6.
1962 Pterocarya paradisiaca (Unger) Ilinskaya, p. 104.

Material. KRAM-P 250: 147/I. cf. Pterocarya paradisiaca: 54/II, 149/II.

Description. Macromorphology. Oblongate leaflet, 3.8 cm long and 1.3 cm wide. Leaflet margin simple serrate, apical side of teeth
concave, basal side convex, straight, teeth apex acute. Branches of secondary veins terminate at the teeth apex. Venation pinnate, primary vein straight, and of moderate thickness. Secondary venation semicraspedodromous. Secondary veins, preserved in up to 7 pairs, depart the primary vein at intervals of 0.35–0.55 cm and form an angle of 60–90° with the primary vein. Secondary veins are upwardly curved and interconnected in loops. Intersecondary veins rarely present. Tertiary venation weakly percurrent or composite intersecondary.

**Micromorphology.** Abaxial epidermis consists of irregularly-shaped cells with rounded to slightly undulate cell walls, 25–37 μm across. Stomata probably anomocytic, wide elliptic, about 20 μm across. Outer stomatal ledge aperture spindle-shaped, about 12 μm long and 7 μm wide. On the abaxial epidermis glandular, peltate trichomes (scales), 100–125 μm in diameter were found. These trichomes have unicellular, elliptic bases, 12–17 μm in diameter. Also one unicellular, globose trichome with unicellular base was found.

**Remarks.** The venation pattern of leaflets and characteristic peltate trichomes with unicellular bases point to species *Pterocarya paradisiaca* (Unger) Ilinskaya. *Carya serrifolia* (Goeppert) Kräusel differs in branched secondary veins having craspedodromous endings (comp. Hummel 1983, Manchester 1987). Two other leaflets (250/54/II and 250/149/II) only presumably belong to *Pterocarya paradisiaca* as they are badly preserved.

**Modern equivalents.** *Pterocarya paradisiaca* is similar to the contemporary *P. fraxinifolia* Spach (Ilinskaya 1968, Knobloch 1969), distributed in the Euxine-Hyrcanian area where it grows along river and stream valleys in riparian forests (Boratyński & Boratyńska 1975, Akhani & Salimian 2003).

**Ecology.** Arctotertiary, warm temperate element. *Pterocarya paradisiaca* is a frequent component of riparian communities in the Palaeogene and Neogene of Eurasia.

**Occurrence.** Common in azonal vegetation of Neogene of Europe, known from the Late Oligocene to the Late Pliocene (Zastawniak et al. 1996). In the territory of Poland found from the Early Miocene to Pliocene (see Worbiec et al. 2008).

Lythraceae Jaume Saint-Hilaire, nom. cons.

**Trapa L.**

**Trapa sp.**

Pl. 3, fig. 5

**Material.** KRAM-P 250: 68/II.

**Description.** Only one small, characteristic fragment of fruit from *Trapa* – barbed spine (also named harpoon) was preserved.

**Remarks.** *Trapa* is an arctotertiary, warm temperate element, a typical water plant. Fruits of several taxa of *Trapa* are common in the Neogene floras of Europe (Wójcicki & Zastawniak 2003).

**Malvaceae Jussieu, nom. cons.**

**? Helicteroideae Griseb.**

**Laria G. Worobiec & Kvaček**

**Laria rueminiana** (Heer) G. Worobiec & Kvaček

Fig. 4: 1, 4–5, Pl. 3., figs 3a–4, Pl. 7, figs 1a–4

1859 *Ficus rueminiana* Heer, p. 183, pl. 152, figs. 11, 12.

1859 *Ficus truncata* Heer, p. 183, pl. 152, fig. 15.

2010 *Laria rueminiana* (Heer) G. Worobiec & Kvaček; Worobiec et al., pp. 903–907, Fig. 6A–6J, Fig. 7A–7H, Fig. 9B.


**Description.** Macromorphology. Only leaf fragments, up to 6.0 cm long and up to 5.7 cm wide. Leaves entire-margined, leaf base truncate to cordate. Petoioles elongated, often with well developed pulvinus. Venation palmate: actinodromous, basal. Primary vein branched into 5 (usually) or 6 (rarely) branches. Secondary venation brochidodromous. Tertiary venation percurrent. About 5 tertiary veins occur per 1 cm of secondary vein or branches of primary vein. Higher-order venation more or less orthogonal reticulate. Areoles well developed, about 0.3 mm across. Veinlets absent or simple. Marginal ultimate venation looped.

**Micromorphology.** Adaxial epidermis composed of isodiametric or slightly elongated
cells, (20) 25–45 (50) μm across. Cell walls in some leaves mostly straight or rounded, in others undulate. Cuticle of adaxial epidermis in some leaves with distinct ornamentation in form of striae while in other leaves without ornamentation. Abaxial epidermis composed of cells 25–42 μm across, anticlinal cell walls usually almost invisible, rounded or straight. Lea-

f ves hypostomatic. Stomata probably anomocytic, elliptic and of variable size, 15–32 μm in diameter, cell walls of guard cells mostly invisible. Some stomata are surrounded by cuti-

cular striations perpendicular to the stomatal pore. Outer stomatal ledge aperture usually spindle-shaped, rarely narrow elliptic, and of very variable size, 11–22 μm long and 4–10 μm wide. On the abaxial epidermis two types of trichomes were found. Most numerous were glandular, clavate-like trichomes, composed of about 4 cells (usually fragmentarily preserved), up to about 52 μm long and 12–20 μm wide with unicellular base, 12–17 (usually 15) μm in diameter. There was also found one, simple, ensiform, unicellular trichome, 57 μm long.

Remarks. The leaves under discussion, both macro- and micromorphologically corre-

spond to the leaf morphotaxon Laria rueminiana (Heer) G. Worobiec & Kvaček that was described as a new combination on the basis of material from the Polish Neogene localities of Bełchatów Lignite Mine and Ruja near Legnica (Worobiec et al. 2010).

Modern equivalents. Fossil Laria rueminiana could be compared with many extant members of Malvaceae s.l. (Worobiec et al. 2010). However, results of investigations on fossil fruits of Reevesia hurnikii Kvaček and associated plant organs (leaves of “Ficus” truncata = Laria rueminiana and winged seeds of Saportaspermum sp.) found in the Early Miocene flora from the Most Basin in North Bohemia, Czech Republic (Kvaček 2006) suggest close relationship between Reevesia and Laria rueminiana. Micromorphologically most of extant Reevesia species differ from Laria rueminiana in dominance of brachyparacytic stomatal complex and in low variability of stoma-

tal size (Worobiec et al. 2010).

Ecology. Palaeotropical, subtropical element. Extant Reevesia, which is considered to be a relative to Laria rueminiana, exists over an area of paratropical to subtropical evergreen forests of the eastern Himalayas and south-

eastern Asia (Kvaček 2006). In the earlier inves-


tigation of Belchatów Lignite Mine and Ruja lignite deposit, Laria rueminiana was found within hygrophilous, mainly swampy and riparian vegetation (Worobiec 2003, Worobiec et al. 2008). However, the assemblage under discussion from Belchatów points to be riparian and mesophytic similarly as vegetation of the North Bohemian Basin locality Čermnýk where numerous leaves of Laria rueminiana were found (Bůžek 1971). Besides, in the Bilina Mine (Czech Republic) leaves of Laria ruem-

iniana were found in assemblages representing vegetation of drier riparian forests on fertile alluvial soils (Parrotia-Ulms association of Kvaček & Boulter in Boulter et al. 1993). There-

fore it can be concluded that Laria rueminiana should be considered as a component of mesophytic to riparian vegetation.

Occurrence. Rare element of the Neogene floras of Europe, in the territory of Poland infrequently found in Middle to Late Miocene assemblages (Worobiec et al. 2010).

Helicteroideae Griseb.

Reevesia Lindl.

Reevesia hurnikii Kvaček

Pl. 4, figs 2a–2b


Material. KRAM-P 250: 77/I.

Description. Coalified detached capsule valve with strongly thickened dorsal edge (see Kvaček 2006), 2.5 cm long and 1.0 cm high.

Remarks. The detached capsule valve is identical with impressions described under the name of Reevesia hurnikii Kvaček by Kvaček (2006) from the Bilina Mine. Kvaček (op. cit.) suggested that fruit valves of Reevesia hurnikii judging to the character of impressions should be woody, and it is confirmed in the material from Belchatów. Of a special interest is co-


occurrence of leaves of Laria rueminiana (Heer) G. Worobiec & Kvaček and fruit valves of Reevesia hurnikii Kvaček in the investigated locality from Belchatów. They are accompanied there by relatively numerous pollen grains of genus Reevesiapollis Krutzsch (Table 1). Only in the
Bílina Mine were found leaves of *Laria rueminianna* and fruit valves of *Reevesia hurnikii* with rather scarce pollen material. Co-occurrence of pollen grains of *Reevesiapollis* with *Laria rueminianna* and *Reevesia hurnikii* is another evidence in support of the idea that leaves of *Laria rueminianna* belong to a fossil representative of the genus *Reevesia*. Moreover, it is very probable that leaves of *Laria rueminianna*, fruit valves of *Reevesia hurnikii* and pollen grains of *Reevesiapollis* represent organs of the same plant.

**Modern equivalents.** The fossil valves of *Reevesia hurnikii* correspond in respect of size and shape to those of several extant species of genus *Reevesia*, e.g. *Reevesia pubescens* Mast. or *R. thyrsoidea* Lindl. (Kvaček 2006).

**Ecology.** Palaeotropical, subtropical element. Extant representatives of *Reevesia* exists over an area of paratropical to subtropical evergreen forests of eastern Himalayas and southeastern Asia (Kvaček 2006). Similarly as in case of co-occurring leaf morphospecies *Laria rueminianna*, *Reevesia hurnikii* should be considered as an element of mesophytic to riparian vegetation.

**Occurrence.** Taxon new for the fossil floras of Poland. Previously found only in the Early Miocene of the Czech Republic (Kvaček 2006).

---

**Salicaceae Mirb.**

**Populus L.**

**Populus balsamoides** Goeppert

Pl. 4, figs 3–4b, Pl. 7, fig. 5

1855 *Populus balsamoides* Goeppert, p. 23, pl. 15, figs. 5–6.

**Material.** KRAM-P 250: 102/I and 112/I (counterparts), 120/I and 126/I (counterparts), 135/III. *Populus cf. balsamoides*: 42/I and 43/I (counterparts), 59/I, 101/IV.

**Description.** Macromorphology. Leaf fragments up to 6.5 cm long, and about 5.0 cm wide. Leaf apex attenuate, base rounded. Margin simple serrate, teeth 2–4 mm long, apical side of teeth very short, concave, basal side long, convex or acuminate, teeth apex rounded. Veins terminate in sinus between teeth. Venation pinnate, primary vein straight and stout. Secondary venation semicraspedodromous, secondaries distributed at intervals of 0.4 cm near leaf base and up to 1.0 cm in the middle of the lamina. Secondary veins upwardly curved diverging from midvein at an angle of 50–60°, branching towards leaf margin forming agrophic (pectinal) veins. Tertiary veins percurrent or composite intersecondary, forms an angle of 90–100° with the primary vein; ca. 8 tertiary veins occur per 1 cm of secondary vein.

**Micromorphology.** Adaxial epidermis consisting of polygonal cells with straight walls, 21–30 μm across. Abaxial epidermal cells 20–35 μm across, cell wall rounded or straight. Leaves probably hypostomatic. Stomatia elliptic to elliptic-elongated, varying in size, 15–25 μm long and 10–15 μm wide. Outer stomatal ledge aperture spindle-shaped, ledges strongly cutinised, 15–20 μm long and 4–9 μm wide.

**Remarks.** Macromorphology of the discussed leaves (shape, network of venation, type and shape of teeth) undoubtedly points to the genus *Populus* and the species *Populus balsamoides*. On the shape of teeth this taxon differs markedly from another fossil poplar, *Populus populina* (Brongniart) Knobloch.

**Modern equivalents.** Extant leaves of *Populus balsamifera* L. are morphologically comparable to *P. balsamoides*. *Populus balsamifera* is North American member of the section *Tacamahaca* Spach, component of riparian forests (Hantke 1954, Krüssmann 1976, Belz & Mosbrugger 1994). Poplars from the section *Tacamahaca* are indigenous to Eastern Asia and North America (Teodoridis et al. 2009). Contrary to this opinion, some of authors compare *Populus balsamoides* with recent Chinese *Populus lasiocarpa* Oliv. from section *Leucoides* Spach. (Ilinskaya 1968, Zastawniak 1972).

**Ecology.** Arctotertiary, temperate element. Woody species of azonal vegetation of riparian forests (Belz & Mosbrugger 1994).

**Occurrence.** Common in the Neogene floras of Europe. In Poland it is found from the Middle Miocene to Pliocene (Zastawniak 1972, Stuchlik et al. 1990, Krajewska 1998).

**Populus populina** (Brongniart) Knobloch

Fig. 4: 7, Pl. 4, figs 7a–7b

1822 *Phyllites populina* Brongniart, p. 237, pl. 14, fig. 4.
1836 *Populus latior* A. Br.; A. Braun in Buckland, p. 512.

**Material.** KRAM-P 250: 118/I.

**Description.** Macromorphology. Fragment of a single leaf, 5.5 x 5.3 cm. Leaf margin simple serrate. Apical side of teeth concave, very rarely straight, basal side convex, very rarely straight, teeth apex acute, sometimes attenuate. Venation pinnate, primary vein bent and of moderate thickness. Secondary venation craspedodromous, secondaries run off primary vein at intervals of 1.2–1.5 cm, diverging from midvein at an angle of 30–50°. Secondaries curved upwards, strongly branched, intersecondary veins present. Tertiary venation weakly percurrent, irregular. Higher-order venation random reticulate.

**Remarks.** Leaf margin with straight or hook-shaped teeth with acute apex are characteristic of fossil poplar *Populus populina* (Brongniart) Knobloch.

**Modern equivalents.** In the respect of macromorphological and micromorphological features to the *Populus populina* are related two extant poplar species, *Populus alba* L. and *Populus tremula* L. from the section Leuce Duby (Knobloch & Kvaček 1976).

**Ecology.** Arctotertiary, temperate element. Component of bottomland vegetation of riparian forests (Belz & Mosbrugger 1994).

**Occurrence.** Neogene floras of Europe. In Poland it occurs from the Middle Miocene to the Pliocene (see Worobiec et al. 2008).

*Populus sp.*


**Description.** Macromorphology. Fragments of sometimes rather large leaves (preserved remains measure up to 8.0 cm). Leaf apex attenuate, base rounded. Leaf margin simple serrate. Apical side of teeth concave, basal side more or less acuminate, teeth apex rounded, hook-shaped. Venation pinnate. Secondary venation craspedodromous, secondary veins diverging from midvein at an angle of 90–100°, secondaries strongly curved upwards. Intersecondary veins present.

**Micromorphology.** Two very poor cuticular slides from specimens 250/71 and 250/123 were obtained. Walls of cells of adaxial epidermis straight. Stomata elliptic. Outer stomatal ledge aperture spindle-shaped, 12–19 μm long and 4–9 μm wide. There were also found (specimen 250/71) numerous unicellular trichome bases on the small fragments of the cuticle from unassigned laminar side.

**Remarks.** The fossil leaf fragments clearly belong to the genus *Populus*, but are too poorly preserved to be assigned to a particular species. Cuticles features observed are very similar to those of *Populus balsamoides* from the same locality.

*Salix L.*

**Salix varians** Goeppert sensu lato

Fig. 4: 6, Pl. 7, fig. 6, Pl. 8, fig. 1

1855 *Salix varians* Goeppert, p. 26, pl. 19, figs 17, 18, pl. 20, fig. 1.

**Material.** KRAM-P 250: 150/II.

**Description.** Macromorphology. Fragment of single, elongated leaf, 3.5 cm long, ca 1.0 cm wide. Leaf apex probably attenuate. Leaf margin simple serrate, teeth rather small, 1.0–1.5 mm long. Apical side of teeth concave, basal side convex. Teeth apex acute, hook-shaped. Secondary vein branches terminate in sinuses between teeth. Venation pinnate, primary vein straight, of moderate thickness. Secondary venation semicraspedodromous, secondaries distributed at intervals of 0.6–0.7 cm, diverging from midvein at an angle of ca 45°. Secondaries curving upwards, running almost parallel to the leaf margin. Intersecondary veins present. Tertiary veins composite intersecondary or percurrent, forming angle of about 100° with primary vein, 10–15 tertiary veins occur per 1 cm.

**Micromorphology.** Cells of abaxial epidermis with straight, rounded, or occasionally undulate walls. Cuticle of abaxial epidermis is covered by epicuticular wax of characteristic micromorphology. Stomata brachyparacytic, narrow elliptic and varying in size, 15–22 μm long and 7–15 μm wide. Outer stomatal ledge aperture narrow spindle-shaped, strongly cutinized, varying in size, 12–19 μm long and
2.5–9.0 µm wide. On the abaxial epidermis, unicellular trichome bases, roundish, 10–15 µm in diameter were found.

Remarks. The macro- and micromorphology of these leaves allowed their identification as *Salix varians* Goeppert. The leaves of *S. varians* show significant morphological variability (Krajewska 1998) and it is possible that this morphospecies includes leaves from more than a single biological species.

Modern equivalents. *Salix varians* is usually compared with the extant Eurasian species of *S. fragilis* L. and *S. triandra* L. (Bůžek 1971, Krajewska 1998).

Ecology. Arctotertiary, temperate element. *Salix varians* was typical species of azonal vegetation of riparian forests (Belz & Mosbrugger 1994).

Occurrence. Common species in the Palaeogene and Neogene floras of Europe, found from the Oligocene to the Pliocene (Krajewska 1998). In the territory of Poland found in the Middle and Late Miocene floras (see Worobiec & Kasiński 2009).

*Sapindaceae* Jussieu, nom. cons.

*Acer* L.

*Acer aegopodifolium* (Goeppert) Baikovskaya ex Ilinskaya

Fig. 5: 1, 3, Pl. 5, figs 3–4b, Pl. 8, figs 2–3b

1855 *Rhus quercifolia* Goeppert, p. 37, pl. 25, figs 6–9.

1855 *Rhus aegopodifolia* Goeppert, p. 37, pl. 25, fig. 10.

1959 *Monopleurophyllum hungaricum* Andreánszky, p. 168, pl. 48, fig. 6, pl. 50, fig. 2, pl. 52, fig. 7, pl. 53, figs 3–4.

1963 *Monopleurophyllum quercifolium* (Goeppert) Kotlaba, p. 63, pl. 5, figs 4, 5.

1965 *Acer aegopodifolium* (Goeppert) Baikovskaya; Shvareva, p. 953.

1968 *Acer aegopodifolium* (Goeppert) Baikovskaya; Ilinskaya, p. 67, pl. 9, figs 12–15, pl. 20, figs 4, 5.

1974 *Acer aegopodifolium* (Goeppert) Baikovskaya ex Ilinskaya; Zhilin, pp. 56, 30.

1988 *Acer quercifolium* (Goeppert) Kovar-Eder, pp. 51–52, pl. 6, figs 14–16.


Description. Macromorphology. Several leaflets of compound leaf, often preserved in original position. Leaflets strongly asymmetric, 2.8–4.0 (4.5) cm long and 2.0–2.5 cm wide, apex acute to rounded, leaf base acute or obtuse. Leaflet margin simple serrate and lobed, teeth rather rare, usually one distinct lobe visible just above leaflet base, apical and basal side of teeth and lobes rounded, apex also rounded. Venation pinnate, midvein straight. Secondary venation mixed craspedodromous, secondary veins irregularly spaced and in the terminal parts characteristically minute bend, strongest branch entering the lobe, some secondaries terminate in teeth and lobe apex, the remaining interconnected in loops. Intersecondary veins present, indistinct. Tertiary and higher order venation more or less random reticulate. Areoles well developed, veinlets present, probably branched. Marginal ultimate venation looped.

Micromorphology. Adaxial epidermis composed of isodiamic or elongated cells, with usually undulate, rarely straight or rounded walls, 32–62 µm across. There are leaflets both with strie ornamentation and without ornamentation of cuticle. Abaxial epidermis composed of isodiamic or elongate cells, with straight or rounded cell walls, 17–27 µm across. Leaflets hypostomatic. Stomata anomocytic, wide elliptic, 20–25 µm long and 17–22 µm wide. Outer stomatal ledge aperture from wide spindle-shaped to elliptic oblongate, 15–17 µm long and 6–8 µm wide.

Remarks. The discussed remains represent
leaflets of compound leaf of the fossil species *Acer aegopodifolium* (Goeppert) Baikovskaya ex Ilinskaya. Baikovskaya (in Shvareva 1965) was the first to identify discussed fossil leaf morphotype as *Acer*. This opinion was confirmed by Ilinskaya (1968). Of the two Goeppert’s species names (“quercifolia” and “aegopodifolia”), name “aegopodifolia” was used to avoid confusion with the species *Rulac quercifolium* Hollick (Ilinskaya 1968, Walther & Zastawniak 2005), described from the Upper Cretaceous deposits of Alaska (Hollick 1930). For the same morphotype of leaf remains as from Belchatów, Kovar-Eder (1988) proposed a new combination, *Acer quercifolium* (Goeppert) Kovar-Eder. However, name *Acer aegopodifolium* (Goeppert) Baikovskaya ex Ilinskaya has priority and *Acer quercifolium* (Goeppert) Kovar-Eder should be treated as synonym. Schmitt and Kvaček (1999) were the first who published data on the micromorphology of the fossil epidermis of the *Acer aegopodifolium* from the Late Miocene deposits of Hambach. Epidermal features of leaflets from Belchatów are much the same. The only found difference is strie ornamentation of the cuticle of leaflets of *Acer aegopodifolium* from Belchatów, yet it is found only in part of leaflets.

**Modern equivalents.** Morphologically *Acer aegopodifolium* could be compared to the extant maples from the section *Trifoliata* Pax (Walther & Zastawniak 2005), especially to *Acer griseum* (Franchet) Pax and *A. trifolium* Komarov.

**Ecology.** Arctotertiary, warm temperate element. Considered to be an accessory species in azonal forest communities (Walther & Zastawniak 2005).

**Occurrence.** *Acer aegopodifolium* is known from the Middle and Upper Miocene of central and eastern Europe (Walther & Zastawniak 2005). From the territory of Poland it is known from Middle Miocene (Młyny, as *Monopleurophyllum quercifolium* (Goeppert) Kotlaba in Zastawniak 1980) and Late Miocene (Sośnica, as *Rhus quercifolia* Goeppert, *R. aegopodofilia* Goeppert in Goeppert 1855, and as *Acer aegopodifolium* in Walther & Zastawniak 2005; Belchatów as *Monopleurophyllum quercifolium* (Goeppert) Kotlaba in Stuchlik et al. 1990; Stare Bystre as *Acer quercifolium* (Goeppert) Kovar-Eder in Worobiec 1994).
**Acer cf. tricuspidatum** Bronn sensu Procházka & Bůžek
Pl. 5, fig. 2

? 1838 *Acer tricuspidatum* Bronn, p. 865, pl. 35, figs 10a, b.
? 1975 *Acer tricuspidatum* Bronn sensu novo, Procházka & Bůžek, p. 24, Figs 2, 3, 4d, 5–13, pl. 22, figs 1–7, pl. 23, figs 1–6, pl. 24, figs 1–4.

**Material.** KRAM-P 250: 26/I.

**Description.** Macromorphology. One badly preserved fragment of lobed and probably entire-margined leaf. Venation palmate, actinodromous. Primary vein branched into 5 branches. Network of tertiary venation typical of genus *Acer*.

**Remarks.** This leaf most probably represents the species *Acer tricuspidatum* Bronn sensu Procházka & Bůžek which is confirmed on the basis of its morphology.

**Modern equivalents.** *Acer tricuspidatum* is classified within section *Rubra* Pax (Walther 1972, Procházka & Bůžek 1975) and is related to the contemporary species *Acer rubrum* L. and *A. saccharinum* L. (Hantke 1954, Kräusel & Weyland 1959). The cuticular micromorphology of extant *A. rubrum*, *A. saccharinum* and *A. hyrcanum* Frisch. & Mey is similar to that of fossil *A. tricuspidatum* (Walther 1972).

**Ecology.** Arctotertiary, temperate element. Usually considered to be a component of swampy vegetation in the Neogene of Central Europe (Walther 1972). As a riparian element it was reported by Kovar-Eder et al. (2001) in the Miocene flora of Oberdorf.

**Occurrence.** *Acer tricuspidatum* is known from Central Europe from the middle Oligocene to the Pliocene, most frequently found in the Miocene floras (Procházka & Bůžek 1975). In the territory of Poland, *Acer tricuspidatum* was a common element of Neogene plant communities (see Worobiec et al. 2008).

**Acer sp.**
Pl. 5, figs 5–6

**Material.** KRAM-P 250: 86/I and 101/II (counterparts).

**Description.** Macromorphology. Maple samara, 2.8–3.2 cm long and 1.2 cm wide (in the widest place). The seed impression about 1 cm long.

**Remarks.** Some morphological similarities could be found with samaras of extant *Acer griseum* (Franchet) Pax, relative of fossil *Acer aegopodifolium* (Goeppert) Baikovskaya ex Ilinskaya, common in the investigated plant assemblage form Belchatów. However, differences found between discussed taxa and scarcity of material (one specimen) excluded reliable assessment to a certain taxon.

**Ulmaceae Mirb.**

**Ulmus L.**

**Ulmus cf. plurinervia** Unger
Fig. 5: 4–5, Pl. 5, fig. 7

? 1847 *Ulmus plurinervia* Unger, p. 95, pl. 25, figs 1–4.

**Material.** KRAM-P 250: 88/I and 89/I (counterparts), 92/I.

**Description.** Macromorphology. Leaf fragments, up to 3.0 cm long and 2.2–2.4 cm wide with obtuse and strongly asymmetric leaf base. Leaf margin serrate, teeth apex moderate acute. Secondary veins terminate in teeth apex. Venation pinnate, primary vein slightly bent, of moderate thickness. Secondary venation simple craspedodromous. Secondary veins, more or less straight, usually opposite, preserved in 10 pairs, run off primary vein at intervals of 0.2–0.3 cm and form an angle of about 60–90° with it.

**Remarks.** Macromorphology of specimens described above is typical for *Ulmus*, e.g. serrate margins with characteristic teeth and dense secondary venation. Relatively small size of leaves and their shape indicate that they could represent a fossil elm *Ulmus plurinervia* Unger.

**Modern equivalents.** East Asiatic species *Ulmus parvifolia* Jacq. and *Ulmus pumila* L. are most similar to *Ulmus plurinervia* Unger (Krajewska 1998).

**Ecology.** Arctotertiary, temperate element. Most probably component of riparian or mesophytic forests.

**Occurrence.** Taxon not very common in the Neogene of Europe. In the territory of Poland
rarely found in the Middle Miocene to Late Miocene floras (see Krajewska 1998).

**Ulmus pyramidalis** Goeppert

Fig. 5: 2, Pl. 5, figs 8–9b

1855 *Ulmus pyramidalis* Goeppert, p. 28, pl. 13, figs 10–12.


**Description.** Macromorphology. Only leaf fragments, up to 5.0 cm long and 2.6–3.6 cm wide with cordate, sometimes rounded and asymmetric leaf base. Petiole, rarely preserved, short (about 0.5 cm) and bent. Leaf margin serrate, teeth characteristic hook-shaped, apical side of teeth concave, basal side convex or acuminate. Teeth apex acute. Secondary veins or their branches terminate in teeth apex. Venation pinnate, primary vein more or less straight, of moderate thickness. Secondary venation simple craspedodromous. Secondary veins rather numerous, up to 15 pairs, run off primary vein at intervals of 0.10–0.15 cm near leaf base and 0.2–0.7 cm (commonly 0.4–0.6 cm) in the middle part of lamina. Secondaries form with primary vein an angle of about 60–90° near leaf base and 30–60° in the middle part of lamina. Secondary veins usually straight or, near the leaf margin, slightly curved upward. Tertiary venation percurrent forming angle of 120–140° with primary vein, 7–12 (commonly 10) tertiary veins occur per 1 cm of secondary vein length. Higher order venation orthogonal reticulate.

**Remarks.** Leaves described above with serrate margins, hook-shaped teeth and dense secondary venation are typical for *Ulmus*. Characteristic features of the above described leaves are rather numerous secondary veins together with elongate shape which are typical for fossil *Ulmus pyramidalis* Goeppert (Hably & Zastawniak 2001).

**Modern equivalents.** *Ulmus pyramidalis* is usually compared with the extant North American species *Ulmus alata* Michx., *Ulmus americana* L., and Chinese *Ulmus castaneifolia* Hemsl. (see Krajewska 1988).

**Ecology.** Arctotertiary, temperate element. Most probably a component of riparian or mesophytic forests. In the Early Miocene (Egerian) floras of Hungary was a dominant element of the riparian vegetation (Hably 1994).

**Occurrence.** Common in the Neogene European floras (Krajewska 1988). In the territory of Poland rarely reported from Early to Late Miocene floras (Krajewska op. cit.).

**Ulmus sp.**


**Remarks.** The characteristic teeth shape, secondary and third order venation of these leaf fragments allow to place them in *Ulmus* L. It is very probable that most of them represent the species *Ulmus pyramidalis* Goeppert, a dominating species of elm in this locality. Nonetheless, the condition of preservation of discussed remains does not allow more precise identification.

**DISCUSSION**

**COMPOSITION OF THE FLORA**

As the result of investigation of 159 deposit samples with plant macroremains, 21 species from genera *Acer*, *Betula*, *Carpinus*, *Dicotylphyllum*, *Fagus*, *Larix*, *Liquidambar*, *Populus*, *Pterocarya*, *Quercus*, *Reevesia*, *Salix*, *Taxodium*, *Trapa*, and *Ulmus* were found. Four new fossil leaf species to the Polish tertiary were reported: *Quercus roburoides* Gaudin, *Reevesia hurnikii* Kvaček, *Dicotylphyllum* sp. 1, and *Dicotylphyllum* sp. 2.

Of special interest is the co-occurrence of morphotaxa from the family Malvaceae s.l. (Tables 1, 2): *Larix rueminiana* (leaves), *Reevesia hurnikii* Kvaček, *Dicotylphyllum* sp. 1, and *Dicotylphyllum* sp. 2.

As the result of investigation of 159 deposit samples with plant macroremains, 21 species from genera *Acer*, *Betula*, *Carpinus*, *Dicotylphyllum*, *Fagus*, *Larix*, *Liquidambar*, *Populus*, *Pterocarya*, *Quercus*, *Reevesia*, *Salix*, *Taxodium*, *Trapa*, and *Ulmus* were found. Four new fossil leaf species to the Polish tertiary were reported: *Quercus roburoides* Gaudin, *Reevesia hurnikii* Kvaček, *Dicotylphyllum* sp. 1, and *Dicotylphyllum* sp. 2.

Of special interest is the co-occurrence of morphotaxa from the family Malvaceae s.l. (Tables 1, 2): *Larix rueminiana* (leaves), *Reevesia hurnikii* Kvaček, *Dicotylphyllum* sp. 1, and *Dicotylphyllum* sp. 2.

Of special interest is the co-occurrence of morphotaxa from the family Malvaceae s.l. (Tables 1, 2): *Larix rueminiana* (leaves), *Reevesia hurnikii* Kvaček, *Dicotylphyllum* sp. 1, and *Dicotylphyllum* sp. 2.

Of special interest is the co-occurrence of morphotaxa from the family Malvaceae s.l. (Tables 1, 2): *Larix rueminiana* (leaves), *Reevesia hurnikii* Kvaček, *Dicotylphyllum* sp. 1, and *Dicotylphyllum* sp. 2.

Of special interest is the co-occurrence of morphotaxa from the family Malvaceae s.l. (Tables 1, 2): *Larix rueminiana* (leaves), *Reevesia hurnikii* Kvaček, *Dicotylphyllum* sp. 1, and *Dicotylphyllum* sp. 2.

Of special interest is the co-occurrence of morphotaxa from the family Malvaceae s.l. (Tables 1, 2): *Larix rueminiana* (leaves), *Reevesia hurnikii* Kvaček, *Dicotylphyllum* sp. 1, and *Dicotylphyllum* sp. 2.

<table>
<thead>
<tr>
<th>Taxon</th>
<th>Botanical affinity</th>
<th>Element</th>
<th>Sample 1</th>
<th>Sample 2</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>SPORES</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baculatisporites primarius + Rugulatisporites sp.</td>
<td>Osmundaceae</td>
<td>P/A</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>Cryptogrammasporis magnoides (Krutzsch)</td>
<td>Pteridaceae: Cryptogramma</td>
<td>A1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Laevisigatosporites spp.</td>
<td>Polypodiaceae, Davalliacae</td>
<td>P/A</td>
<td>7</td>
<td>2</td>
</tr>
<tr>
<td>Leiotritelles sp.</td>
<td>Lygodiaceae</td>
<td>P</td>
<td>3</td>
<td>1</td>
</tr>
<tr>
<td>Retiritelles sp.</td>
<td>Lycopodiaceae: Lycopodium</td>
<td>A</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>other spores</td>
<td>ferns</td>
<td>unknown</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td><strong>GYMNOSPERMS</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Abiespollenites sp.</td>
<td>Pinaceae: Abies</td>
<td>A</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Cathayapollis spp.; Pinuspollenites sp.</td>
<td>Pinaceae: Cathaya; Pinus haplohyton type</td>
<td>A1</td>
<td>11</td>
<td>7</td>
</tr>
<tr>
<td>Inaperturopollenites spp.</td>
<td>Cupressaceae: Taxodium, Glyptostrobus</td>
<td>P2/A1</td>
<td>19</td>
<td>33</td>
</tr>
<tr>
<td>Piceapollis spp.</td>
<td>Pinaceae: Picea</td>
<td>A</td>
<td>1</td>
<td>5</td>
</tr>
<tr>
<td>Pinuspollenites labadcus (Potonié) Raatz</td>
<td>Pinaceae: Pinus; Pinus sylvestris type</td>
<td>A</td>
<td>33</td>
<td>29</td>
</tr>
<tr>
<td>Sciadopityspollenites sp.</td>
<td>Sciadopityaceae: Sciadopitys</td>
<td>A1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Sequoia pollenites sp.</td>
<td>Cupressaceae: Sequoia, Sequoiadendron, Metasequoia</td>
<td>A1</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>Zonalapollenites spp.</td>
<td>Pinaceae: Tsuga</td>
<td>A</td>
<td>6</td>
<td>4</td>
</tr>
<tr>
<td><strong>ANGIOSPERMS</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aceripollenites striatus (Pflug) Thiele-Pfeiffer</td>
<td>Sapindaceae: Acer</td>
<td>A1</td>
<td>5</td>
<td>8</td>
</tr>
<tr>
<td>Alninpollenites verus Potonié</td>
<td>Betulaceae: Alnus</td>
<td>P2/A</td>
<td>23</td>
<td>15</td>
</tr>
<tr>
<td>Caprifoliipites sp.</td>
<td>Adoxaceae: Sambucus, Viburnum</td>
<td>P/A1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Carpinites carpoides (Pflug) Nagy</td>
<td>Betulaceae: Carpinus</td>
<td>P2/A1</td>
<td>20</td>
<td>33</td>
</tr>
<tr>
<td>Caryapollenites simplex (Potonié) Raatz</td>
<td>Juglandaceae: Carya</td>
<td>A1</td>
<td>4</td>
<td>2</td>
</tr>
<tr>
<td>Cupuliferapollenites oviformis (Potonié) Potonié</td>
<td>Fagaceae: Castanea, Castanopsis, Lithocarpus</td>
<td>P2/A1</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>Celtipollenites sp.</td>
<td>Ulmaceae: Celtis</td>
<td>P/A1</td>
<td>4</td>
<td>3</td>
</tr>
<tr>
<td>Cyperaceae pollenites neogenicus Krutzsch</td>
<td>Cyperaceae</td>
<td>P/A</td>
<td>6</td>
<td>10</td>
</tr>
<tr>
<td>Eriicipites sp.</td>
<td>Ericaceae</td>
<td>A</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>Faguspollenites sp.</td>
<td>Fagaceae: Fagus</td>
<td>A</td>
<td>31</td>
<td>62</td>
</tr>
<tr>
<td>Graminidites spp.</td>
<td>Poaceae: Pooidae</td>
<td>P/A</td>
<td>3</td>
<td>5</td>
</tr>
<tr>
<td>Illepollenites margaritatus (Potonié) Raatz</td>
<td>Aquifoliaceae: Ilex</td>
<td>P/A1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Intratriporopollenites sp.</td>
<td>Malvaceae: Tilioidae</td>
<td>A1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Juglanspollenites verus Raatz</td>
<td>Juglandaceae: Juglans</td>
<td>A1</td>
<td>1</td>
<td>6</td>
</tr>
<tr>
<td>Myricipites sp.</td>
<td>Myricaceae</td>
<td>P2/A1</td>
<td>4</td>
<td>1</td>
</tr>
<tr>
<td>Nyssapollenites sp.</td>
<td>Cornaceae: Nyssa</td>
<td>P2/A1</td>
<td>10</td>
<td>3</td>
</tr>
<tr>
<td>Oleoidearumpollenites sp.</td>
<td>Oleaceae: ?Fraxinus</td>
<td>A1</td>
<td>3</td>
<td>2</td>
</tr>
<tr>
<td>Periporopollenites stigmaticus (Potonié) Thomson &amp; Pflug</td>
<td>Altingiaceae: Liquidambar</td>
<td>A1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Polytrapollenites stellatus (Potonié) Pflug</td>
<td>Juglandaceae: Pterocarya</td>
<td>A1</td>
<td>12</td>
<td>10</td>
</tr>
<tr>
<td>Potamogetonaccidites sp.</td>
<td>Potamogetonaceae: Potamogeton</td>
<td>P/A</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>?Quercides henrici (Potonié) Potonié, Thomson &amp; Thiergart</td>
<td>Fagaceae: Quercoideae</td>
<td>P/A1</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>Quercopollenites sp.</td>
<td>Fagaceae: Quercus</td>
<td>A1</td>
<td>21</td>
<td>31</td>
</tr>
<tr>
<td>Reevesiapollis sp.</td>
<td>Malvaceae: Reevesia</td>
<td>P</td>
<td>2</td>
<td>7</td>
</tr>
<tr>
<td>Salizipollenites sp.</td>
<td>Salicaceae: Salix</td>
<td>A2</td>
<td>1</td>
<td>4</td>
</tr>
<tr>
<td>Sparganiaceae pollenites sp.</td>
<td>Sparganiaceae, Typhaceae</td>
<td>P/A</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>Tricolporopollenites exactus (Potonié) Grabowska</td>
<td>Cyrtalceae, Clethraceae</td>
<td>P2/A1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>?Tricolporopollenites pseudocingulum (Potonié) Thomson &amp; Pflug</td>
<td>?Styracaceae</td>
<td>P/A1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Tricolpapollenites sp.</td>
<td>Rosaceae</td>
<td>P/A</td>
<td>1</td>
<td></td>
</tr>
</tbody>
</table>
along with Reevesiapollis pollen which are rather abundant (Table 1). This pollen type was also present in one of the cuticular slides of Laria rueminiana, occurring in clumps. The co-occurrence of the three malvalean morpho-taxa in Bełchatów suggests that they might represent vegetative and reproductive organs of a single biological species and undoubtedly confirming the presence of Reevesia in the fossil flora from the Late Miocene of Bełchatów Mine.

**TAPHONOMY**

Plant macroremains were preserved as dispersed in fine-grained deposits. The structure of deposits suggests that the assemblage was formed in water reservoir with stagnant or slow-flowing water, probably oxbow lake of meandering river or abandoned channel of braided river. Presence of shallow, stagnant waters is confirmed by occurrence of Trapa macroremain and pollen grains of Potamogeton, plants typical of this environment (Table 1). The rate of sedimentation was especially high during periodical floods. Swelled rivers delivered detritic material that had immediately buried plant remains accumulated on the reservoir bottom and prevented extensive decay or complete decomposition of them. The discussed fossil flora was parautochtonous with considerable admixture of allochtonous taxa (comp. Gastaldo et al. 1996). It is confirmed by the mixture of taxa characteristic of azonal (riparian) and zonal (mesophytic) vegetation. Such a mixture of taxa could indicate that the sedimentary reservoir was situated at floodplain covered by riparian forests near elevated and drier river terraces or levee being typically covered with mesophytic vegetation. The same mixture of vegetation could be inferred from the results of palynological investigations.

**PALYNOLOGY OF PLANT ASSEMBLAGE**

Both studied samples yielded rich and well-preserved sporomorph assemblages suitable for detailed pollen analysis. The palynoflora is composed of 12 gymnosperm pollen, 36 angiosperm pollen, and 8 spore morpho-taxa (Table 1). The identified sporomorph taxa represent plants that probably occurred in the following plant communities:

**Azonal vegetation communities:**

Swamp forest (composed mainly of Taxodioideae, including Taxodium and Glyptostrobus, as well as Nyssa and Alnus)

Riparian forest (Pterocarya, Ulmus, Acer, Juglans, Salix, Alnus, Celtis, Carya, Fraxinus, Reevesia, and others)

Reeds and surrounding vegetation (mainly Cyperaceae, Poaceae, and Sparganium)

Aquatic plants (Potamogeton)

**Zonal vegetation community:**

Mesophytic forest (composed mainly of deciduous trees Fagus, Quercus, Carpinus, Betula, Ulmus, Castanea, Juglans, Acer, and coniferous trees Pinus, Cathaya, Tsuga, Picea, and others).

The identified sporomorph taxa have been classified to an appropriate palaeofloristic element (see Table 1), mainly on the basis of Atlas of pollen and spores of the Polish Neogene (Stuchlik et al. 2001, 2002, 2009, in press), and the checklist of selected pollen and spore taxa from the Neogene deposits proposed by Ziembińska-Tworzydło et al. (1994). In the studied material the following palaeofloristic elements have been distinguished: palaeotropical (P), including subtropical (P2); arcto-tertiary (A), including: warm-temperate (A1) and temperate (A2), as well as cosmopolitan (PA).

The composition of sporomorph associations from the studied samples shows an apparent

<table>
<thead>
<tr>
<th>Taxon</th>
<th>Botanical affinity</th>
<th>Element</th>
<th>Sample 1</th>
<th>Sample 2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Triporopollenites coryloides Pflug</td>
<td>Betulaceae: Corylus</td>
<td>A</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>Trivestibulopollenites betuloides Pflug</td>
<td>Betulaceae: Betula</td>
<td>A</td>
<td>52 58</td>
<td></td>
</tr>
<tr>
<td>Umbelliferopollenites sp.</td>
<td>Apiaceae</td>
<td>A2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Vitispollenites sp.</td>
<td>Vitaceae: Vitis</td>
<td>A1</td>
<td>3 2</td>
<td></td>
</tr>
<tr>
<td><strong>VARIA</strong> pollen and spores indet.</td>
<td>unknown</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Sum of sporomorphs</strong></td>
<td></td>
<td></td>
<td>342 406</td>
<td></td>
</tr>
</tbody>
</table>

Table 1. Continued
dominance of arctotertiary (including warm-temperate and temperate) and cosmopolitan palaeofloristic elements (Table 1). Palaeotropical elements are represented by a few taxa only (mainly subtropical), all occurring as rare specimens. A few spores of *Leiotritetes* sp. and pollen grains of *Cupuliferoipollenites outiformis*, *Ilexpollenites margaritatus*, *Reeveriapolles* sp., *Tricolporopollenites exactus*, as well as probably *Quercoidites henrici* and *Tricolporopollenites pseudocingulum*, have been encountered.

**CHARACTERISTICS OF PALEOVEGETATION**

On the basis of investigations on plant macroremains and palynological analyses, reconstruction of the local vegetation of the discussed Late Miocene locality from Belchatów Lignite Mine has been performed. The area surrounding the sedimentary reservoir in which formed the fossil assemblage was covered by riparian (azonal) and mesophytic (zonal) forests.

Riparian vegetation was composed of *Acer cf. tricuspidatum*, *Larix rueminiana*, *Liquidambar europaea*, *Populus balsamoides*, *Populus populina*, *Populus sp.*, *Pterocarya paradipliaca*, *Reevesia hurnikii*, *Salix sp.*, and *Salix varians* sensu lato. Palynological analysis supplemented the list of riparian taxa of discussed assemblage with genera *Alnus*, *Carya*, *Fraxinus*, *Sequoia*, and *Vitis*. Riparian forests of this type occurred in the European tertiary from the middle Palaeocene to the end of Pliocene (Mai 1995). Recent equivalents to the Late Miocene riparian vegetation from Belchatów exist in south-eastern North America (Knapp 1965, Barnes 1991) as well as in the Euxine-Hyrcanian area (Anatolia, Colchis, Talysh, northern China (Wang 1965, Barnes 1991), northern China (Wang 1961), and Euxine-Hyrcanian area (Akhani et al. 2010).

Among macroremains swamp forests taxa are represented only by *Taxodium dubium*. Pollen analysis revealed also scarcity of swamp elements in the fossil assemblage, confirming only the presence of Taxodioidae (*Taxodium*/Glyptostrobus) and *Nyssa*. These trees occurred there as remnants of swamp forests of the *Glyptostrobus-Alnus-Bytneriophyllum* association (Mai 1995) which existed in Europe from the Upper Oligocene to the Pliocene, dominating in the Miocene of the Polish Lowlands area.

Presence of harpoon from a *Trapa* fruit and pollen grains of *Potamogeton* and Sparaganiaceae/Typhaceae suggests that the fossil plant assemblage accumulated in a water reservoir surrounded by reed (Sparaganiaceae/ Typhaceae) vegetation. The water surface was most likely covered by floating leaves of *Trapa* and *Potamogeton*. The occurrence of *Trapa* and *Potamogeton* suggest stagnant or only slow running water pointing to the existence of an oxbow lake.

**RECONSTRUCTION OF PALEOCoclIMATE**

Most of the fossil taxa of plant macroremains represent the arctotertiary geofloristic element (Mai 1995). Extant taxa that are closely related to the fossil species are presently thriving under warm temperate climate in the northern hemisphere. Taxa representing palaeotropical element like *Reevesia* are rare both in macro- and microremains. Absence of thermophilous taxa such as Lauraceae or palms, the rarity of palaeotropical species, and the dominance of arctotertiary element indicates a warm temperate climate during the accumulation of the fossil plant assemblage.
The palaeoclimate may have been comparable to contemporary climates of the area of Euxine-Hyrcanian forests. In riparian forests of this region _Pterocarya fraxinifolia_ C.A. Mey occurs that is comparable to the fossil _Pterocarya paradisiaca_ (Unger) Ilinskaya from Belchatów. The Euxine-Hyrcanian area has favourable climatic conditions for vegetation, and according to Akhani et al. (2010) it endures high amounts of annual precipitation that is evenly distributed throughout the year (very short or absent dry season). The average minimum temperature for the coldest month is higher than 0°C. According to Tutayuk (1975) the climatic values of this area are as follows: mean annual temperatures around +14°C, mean temperature of the coolest month above 0°C, mean annual precipitation more than 1000 mm.

The palaeoclimate as reconstructed from the fossil assemblage from Belchatów could also be compared to that of south-eastern North America, especially along the Atlantic coastline. For instance, _Acer rubrum_ var. _drummondii_ (Hook & Arnott) Sarg. occurs in this area and is believed to be closely related to the fossil _Acer tricuspidatum_. The climate of this region is characterised by a mean annual temperature varying between +14 and +19°C, the mean temperature of the coolest month ≥ +5°C, and high annual precipitation, up to 1200–1500 mm (Knapp 1965, Barnes 1991).

AGE OF THE PLANT ASSEMBLAGE

From the geological point of view, the discussed fossil assemblage from the Belchatów Lignite Mine is situated at the lower part of clayey-sandy unit which age is estimated as Late Miocene (see Geology). This is in accordance with results of our palaeobotanical investigations. Early Miocene age is excluded due

Table 2. Taxa shared between the KRAM-P 250 plant assemblage and selected leaf floras from the Neogene of Poland. (+) – taxon given under another name

<table>
<thead>
<tr>
<th>Taxon</th>
<th>Middle Miocene</th>
<th>Late Miocene</th>
<th>Pliocene</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Acer cf. tricuspidatum</em></td>
<td>(+)</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Acer aegopodifolium</em></td>
<td></td>
<td>(+) (+)</td>
<td>(+)</td>
</tr>
<tr>
<td><em>Betula pirolatypetera</em></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Betula subpubescens</em></td>
<td>(+)</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Carpinus grandis</em></td>
<td>(+)</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Fagus silesiaca</em></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Larix rueminiana</em></td>
<td></td>
<td>+</td>
<td>?</td>
</tr>
<tr>
<td><em>Liquidambar europaea</em></td>
<td>(+)</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Populus balsamoides</em></td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Populus populina</em></td>
<td>(+)</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Pterocarya paradisiaca</em></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Quercus pseudocastanea</em></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Quercus roburoides</em></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Rewisia harnikii</em></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Salix varians</em></td>
<td>(+)</td>
<td>+</td>
<td>+ ?</td>
</tr>
<tr>
<td><em>Taxodium dubium</em></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Trapa sp.</em></td>
<td></td>
<td>(+)</td>
<td>+</td>
</tr>
<tr>
<td><em>Ulmus cf. plurinervia</em></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Ulmus pyramidalis</em></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>
to absence of the members of the family Lauraceae and palms (Areceaceae) and scarcity of palaeotropical elements both in macro- and microremains. Floristic composition of the plant assemblage is generally typical of late Neogene floras of Central Europe. Most of the taxa occur from Miocene to Pliocene, but most frequently in Middle and Late Miocene. Morphotaxa of representative of the genus Reevesia (leaves of Laria rueminiana and fruit capsule valve of Reevesia hurnikii) are restricted to Miocene floras of Europe. Acer aegopodifolium found in the discussed locality from Belchatów, were reported only from deposits older than Pliocene. Quercus roburoides is the only one species typical of Pliocene floras of Europe (van der Burgh 1993). However, rarely it is also found in slightly older deposits, for example of uppermost Miocene to lowermost Pliocene of the Lower Rhine Embayment (Hauptkies Serie, Belz & Mosbrugger 1994). Taking into consideration domination of the warm temperate, deciduous floristic element among plant remains and earlier discussion, we suggest Late Miocene age of plant macroremains.

Moreover, composition of pollen spectra along with low frequency of sporomorphs representing palaeotropical element points to the Late Miocene age of the sediment studied and thereby of the corresponding horizons with plant macroremains. Therefore, the results of palynological investigations definitely confirm the Late Miocene age of this part of clayey-sandy unit of Neogene deposits of the Belchatów Lignite Mine.

**COMPARISON OF THE PLANT ASSEMBLAGE FROM BEŁCHATOW WITH OTHER NEOGENE LEAF FLORAS OF POLAND AND CENTRAL EUROPE**

The macroremains assemblage is taxonomically very similar to other late Neogene leaf floras of Poland (Table 2). Most similar floras are earlier described Late Miocene fossil plant assemblages from the Belchatów Lignite Mine (Stuchlik et al. 1990, Worobiec & Lesiak 1998, Worobiec 2003, Worobiec & Szynkiewicz 2007, Worobiec et al. 2010). Pliocene floras from

<table>
<thead>
<tr>
<th>Taxon</th>
<th>Middle Miocene</th>
<th>Middle Miocene/Late Miocene</th>
<th>Late Miocene</th>
<th>Pliocene</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acer cf. tricuspidatum</td>
<td>(Ferguson 1971)</td>
<td>(+)</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Acer aegopodifolium</td>
<td></td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Betula pioplatyptera</td>
<td></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Betula subpubescens</td>
<td></td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Carpinus grandis</td>
<td></td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Fagus silesiaca</td>
<td></td>
<td>(+)</td>
<td>(+)</td>
<td>(+)</td>
</tr>
<tr>
<td>Laria rueminiana</td>
<td>?</td>
<td>(+)</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Liquidambar europaea</td>
<td></td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Populus balsamoides</td>
<td></td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Populus populina</td>
<td></td>
<td>(+)</td>
<td>+</td>
<td>?</td>
</tr>
<tr>
<td>Pterocarya paradisiaca</td>
<td>?</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Quercus pseudocastanea</td>
<td></td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Reevesia hurnikii</td>
<td></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Salix varians</td>
<td></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Taxodium dubium</td>
<td></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Trapoa sp.</td>
<td></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Ulmus cf. plurinervia</td>
<td></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Ulmus pyramidalis</td>
<td></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Reevesia hurnikii</td>
<td></td>
<td></td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>
Ruszów (Hummel 1983, 1991) and Domański Wierch (Zastawniak 1972) share many taxa with the Bełchatów flora (Table 2). Other localities have less taxa in common (Table 2), but it is usually due to low taxonomical diversity and different sedimentary environments as the other assemblages are mostly derived from swampy communities.

Macrofossil plant assemblages similar to the KRAM-P 250 from Bełchatów have been reported from numerous localities outside Poland (cf. Mai 1995). The taxonomic composition of the Bełchatów assemblage compares well with Middle/Late Miocene leaf floras of central and western Europe (Table 3), e.g. Precarpathians (Shvareva 1983), Achldorf (Knobloch 1986), and Molassezone of Austria (Kovar-Eder 1988). It compares much less with older floras (Table 3), e.g. Kreuzau (Ferguson 1971) and younger floras, e.g. Willershausen (Knobloch 1998).

**CONCLUSIONS**

- As the result of investigation of 159 rock samples with plant macroremains, 21 species, representing 15 genera, belonging to 10 families of gymnosperms and angiosperms have been identified.
- Four new fossil species are reported for the tertiary of Poland: *Quercus roburoides* Gaudin, *Reevesia hurnikii* Kvaček, *Dicotylophyllum* sp. 1, and *Dicotylophyllum* sp. 2.
- Palynological analysis show a mixture of taxa representing riparian (azonal) vegetation and zonal mesophytic forests. Sporomorphs representing azonal swamp forest vegetation are relatively rare. Taxa belonging to arctotertiary, warm temperate element prevail in the pollen spectra. Palaeotropical element is very rare.
- Local vegetation surrounding the sedimentary reservoir was dominated by riparian (azonal) and mesophytic (zonal) forests. Riparian forests were composed of *Acer*, *Larix*, *Liquidambar*, *Populus*, *Pterocarya*, *Reevesia*, *Salix*, and representatives of genera *Alnus*, *Carya*, *Fraxinus*, *Sequoia*, and *Vitis* as indicated by pollen analysis. Zonal vegetation of mesophytic forests from drier areas in the vicinity were composed of angiosperms such as *Acer*, *Betula*, *Carpinus*, *Fagus*, *Quercus*, *Ulmus*, as well as representatives of genera *Castanea*, *Celtis*, *Corylus*, *Juglans*, *Picea*, *Pinus*, and *Tsuga*, probably also *Abies* and *Sciadopitys* as revealed by palynological analysis. Swamp forests taxa were represented only by shoots of *Taxodium* and pollen grains of *Nyssa* and *Taxodioidae*, remnants of *Glyptostrobus-Alnus-Byttnierophyllum* association (Mai 1995). Macroremain of *Trapa* and pollen grains of *Potamogeton* and *Sparganiaceae/Typhaceae* suggest existence of a water reservoir with stagnant or only very slowly moving water.
- Fossil plant taxa found (both macro- and microremains) represent mainly warm temperate and temperate arctotertiary element and indicate warm temperate climate during the period of formation of plant assemblage. Palaeoclimatic condition could be similar to the contemporary climate of the area of Euxine-Hyrcanian forests and/or south-eastern part of North America.
- The stratigraphical position of the assemblage KRAM-P 250 in the profile of Neogene deposits of the Belchatów Lignite Mine and its floristic composition point to the Late Miocene age.

**ACKNOWLEDGEMENTS**

This study was financed by the W. Szafer Institute of Botany, Polish Academy of Sciences in Krakow through the statutory founds. We would like to thank M.Sc. Katarzyna Cywa (Department of Palaeobotany, W. Szafer Institute of Botany, Polish Academy of Sciences in Krakow) for help in taking some of the photographs of plant macrofossils. The authors also thank Prof. dr. Zlatko Kvaček (Charles University, Prague, Czech Republic), and Dr. Friðgeir Grimsson (University of Vienna) for critical reading the manuscript and valuable comments.

**REFERENCES**


APG III. 2009. An update of the Angiosperm Phylogeny Group classification for the orders and families


PLATES

Plate 1

Liquidambar europaea A. Braun

1. Leaf, specimen No. KRAM-P 250/17/I
2. Detail of leaf margin, specimen No. KRAM-P 250/7/I
3. Leaf, specimen No. KRAM-P 250/10/I

Betula plioplatyptera Hummel

4. Detail of leaf margin, specimen No. KRAM-P 250/83/I
6. Leaf, specimen No. KRAM-P 250/13/I

Taxodium dubium (Sternberg) Heer

5. Twig, specimen No. KRAM-P 250/31/I

Betula subpubescens Goeppert emend. Worobiec

7. Leaf, specimen No. KRAM-P 250/74/I

1–3, 5–7: scale bar – 1 cm; 4: scale bar – 1 mm
Plate 2

Betula subpubescens Goeppert emend. Worobiec

1. Leaf, specimen No. KRAM-P 250/133/I
   Carpinus grandis Unger emend. Heer

2. Leaf, specimen No. KRAM-P 250/113/I/A
   4a. Leaf, specimen No. KRAM-P 250/113/I/B
   4b. Detail of leaf margin, specimen No. KRAM-P 250/113/I/B

Fagus silesiaca Walther & Zastawniak

3. Leaf, specimen No. KRAM-P 250/131/I
8. Detail of tooth venation, specimen No. KRAM-P 250/16/I
   Dicotylophyllum sp. 1

5. Leaf, specimen No. KRAM-P 250/104/I
6. Detail of leaf margin venation, specimen No. KRAM-P 250/107/I
   Carpinus grandis Unger sensu Berger

7. Fruit involucre, specimen No. KRAM-P 250/5/I

1, 2, 3, 4a, 5, 7: scale bar – 1 cm; 4b, 6, 8: scale bar – 1 mm
Plate 3

*Quercus roburoides* Gaudin

1a. Leaf, specimen No. KRAM-P 250/21/I
1b. Detail of lobe shape and venation, specimen No. KRAM-P 250/21/I

*Quercus pseudocastanea* Goeppert emend. Walther & Zastawniak

2a. Leaf, specimen No. KRAM-P 250/136/II
2b. Detail of lobe shape and venation, specimen No. KRAM-P 250/136/II

*Laria rueminiana* (Heer) G. Wórhíce & Kvaček

3a. Leaf, specimen No. KRAM-P 250/94/I
3b. Detail of leaf base and petiole with pulvinus, specimen No. KRAM-P 250/94/I
3c. Detail of leaf margin venation, specimen No. KRAM-P 250/94/I
4. Leaf, specimen No. KRAM-P 250/50/I

*Trapa* sp.

5. Barbed spine of fruit, specimen No. KRAM-P 250/68/II

*Pterocarya paradisiaca* (Unger) Ilinskaya

6a. Leaflet, specimen No. KRAM-P 250/147/I
6b. Detail of leaflet margin, specimen No. KRAM-P 250/147/I

1a, 2a, 3a, 3b, 4, 6a: scale bar – 1 cm; 1b, 2b, 5, 6b: scale bar – 1 mm; 3c: scale bar – 200 μm
Plate 4

*Dicotylophyllum* sp. 2

1. Leaf, specimen No. KRAM-P 250/48/I

*Reevesia hurnikii* Kvaček

2a. Detached capsule valve, specimen No. KRAM-P 250/77/I
2b. Detached capsule valve, specimen No. KRAM-P 250/77/I

*Populus balsamoides* Goeppert

3. Leaf, specimen No. KRAM-P 250/112/I
4a. Leaf, specimen No. KRAM-P 250/102/I
4b. Detail of teeth and leaf margin venation, specimen No. KRAM-P 250/102/I

*Populus sp.*

5. Leaf, specimen No. KRAM-P 250/145/I

*Salix varians* Goeppert *sensu lato*

6a. Leaf, specimen No. KRAM-P 250/150/II
6b. Detail of teeth and leaf margin venation, specimen No. KRAM-P 250/150/II

*Populus populina* (Brongniart) Knobloch

7a. Leaf, specimen No. KRAM-P 250/118/I
7b. Detail of teeth and leaf margin venation, specimen No. KRAM-P 250/118/I

1–3, 4b, 5, 6a, 7a: scale bar – 1 cm; 4a, 6b, 7b: scale bar – 1 mm
Plate 5

*Salix* sp.

1. Leaf, specimen No. KRAM-P 250/93/I

   *Acer cf. tricuspidatum* Bronn sensu Procházka & Bůžek

2. Leaf, specimen No. KRAM-P 250/26/I

   *Acer aegopodifolium* (Goeppert) Baikovskaya ex Ilinskaya

3. Fragment of compound leaf, specimen No. KRAM-P 250/83/I
4a. Fragment of compound leaf, specimen No. KRAM-P 250/120/II
4b. Detail of lobe venation, specimen No. KRAM-P 250/120/II

   *Acer* sp.

5. Samara, specimen No. KRAM-P 250/86/I
6. Samara, specimen No. KRAM-P 250/101/II

   *Ulmus cf. plurinervia* Unger

7. Leaf, specimen No. KRAM-P 250/88/I

   *Ulmus pyramidalis* Goeppert

8. Leaf, specimen No. KRAM-P 250/56/I
9a. Leaf, specimen No. KRAM-P 250/33/I
9b. Detail of teeth and leaf margin venation, specimen No. KRAM-P 250/33/I

1–4a, 5–9a: scale bar – 1 cm; 4b, 9b: scale bar – 1 mm
Plate 6

*Taxodium dubium* (Sternberg) Heer

1. Epidermis. Note the oblique arrangement of longer axes of stomata, specimen No. KRAM-P 250/31/I

*Betula subpubescens* Goeppert emend. Worobiec

2. Multicellular base of glandular trichome on abaxial epidermis, specimen No. KRAM-P 250/133/I

*Dicotylophyllum* sp. 1

3a. Abaxial epidermis with stomata, specimen No. KRAM-P 250/99/I
3b. One-celled base of trichomes on abaxial epidermis, specimen No. KRAM-P 250/99/I

*Dicotylophyllum* sp. 2

4. One-celled base of trichomes on abaxial epidermis, specimen No. KRAM-P 250/152/I

*Fagus silesiaca* Walther & Zastawniak

5. Abaxial epidermis with stoma, specimen No. KRAM-P 250/126/III

*Pterocarya paradisiaca* (Unger) Ilinskaya

6a. Peltate glandular trichomes with unicellular base, specimen No. KRAM-P 250/147/I
6b. Detail of peltate glandular trichome, specimen No. KRAM-P 250/147/I

6a: scale bar – 100 μm; 1, 4, 6b: scale bar – 50 μm; 2, 3a, 3b, 5: scale bar – 20 μm
Plate 7

*Laria rueminiana* (Heer) G. Worobiec & Kvaček

1a. Adaxial epidermis, specimen No. KRAM-P 250/80/I
1b. Detail of adaxial epidermis, specimen No. KRAM-P 250/80/I
1c. Abaxial epidermis with stomata, specimen No. KRAM-P 250/80/I
2. Abaxial epidermis with stomata, specimen No. KRAM-P 250/94/I
3. Detail of stomata structure and striations, specimen No. KRAM-P 250/79/I
4. Four-celled glandular clavate trichome, specimen No. KRAM-P 250/54/III

*Populus balsamoides* Goeppert

5. Abaxial epidermis with stomata, specimen No. KRAM-P 250/120/I

*Salix varians* Goeppert *sensu lato*

6. Abaxial epidermis. Note brachyparacytic stomata and cover of epicuticular wax, specimen No. KRAM-P 250/150/II

1a, 1c, 2, 5, 6: scale bar – 50 μm; 1b, 3, 4: scale bar – 20 μm
Plate 8

*Salix varians* Goeppert *sensu lato*

1. Details of brachyparacytic stomata and cover of epicuticular wax, specimen No. KRAM-P 250/150/II

*Acer aegopodifolium* (Goeppert) Baikovskaya ex Ilinskaya

2. Adaxial epidermis, specimen No. KRAM-P 250/47/I

3a. Details of adaxial epidermis. Note striae ornamentation of cuticle, specimen No. KRAM-P 250/4/I

3b. Abaxial epidermis with stomata, specimen No. KRAM-P 250/4/I

2: scale bar – 100 μm; 1, 3a, 3b: scale bar – 20 μm